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Crystal Structure of the P Pilus Rod Subunit PapA

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P pili are important adhesive fibres involved in kidney infection by uropathogenic Escherichia coli strains. P pili are assembled by the conserved chaperone-usher pathway, which involves the PapD chaperone and the PapC usher. During pilus assembly, subunits are incorporated into the growing fiber via the donor–strand exchange (DSE) mechanism, whereby the chaperone’s G1 β-strand that complements the incomplete immunoglobulin-fold of each subunit is displaced by the N-terminal extension (Nte) of an incoming subunit. P pili comprise a helical rod, a tip fibrillum, and an adhesin at the distal end. PapA is the rod subunit and is assembled into a superhelical right-handed structure. Here, we have solved the structure of a ternary complex of PapD bound to PapA through donor–strand complementation, itself bound to another PapA subunit through DSE. This structure provides insight into the structural basis of the DSE reaction involving this important pilus subunit. Using gel filtration chromatography and electron microscopy on a number of PapA Nte mutants, we establish that PapA differs in its mode of assembly compared with other Pap subunits, involving a much larger Nte that encompasses not only the DSE region of the Nte but also the region N-terminal to it.

Introduction

Urinary tract infections, which include infections of the bladder (cystitis) and kidney (pyelonephritis), are some of the most common bacterial infections. These infections are caused mainly by uropathogenic Escherichia coli [1]. Once uropathogenic E. coli is introduced, survival and persistence of these bacteria in the urinary tract require a specific set of virulence factors, including the expression of type P pili. P pili are specifically required for the ability of uropathogenic E. coli to bind Gal-α (1–4)-Gal moieties in human kidney cells and cause pyelonephritis [2,3]. P pili are encoded by the pap gene cluster and are assembled via the highly conserved chaperone-usher pathway, involving the periplasmic immunoglobulin (Ig)-like chaperone PapD and an outer membrane usher PapC [4,5]. P pili consist of six subunits making up a composite fiber with a short tip fibrillum composed of the PapE subunit joined to a more rigid helical rod composed of the PapA subunit [6,7]. PapG is the adhesin at the end of a tip fibrillum; PapK and PapF are adaptor subunits between the PapA rod and the PapE fibrillum and between the PapE fibrillum and the PapG adhesin, respectively; finally, PapH terminates P pilus formation [8,9]. The PapA rod is formed by more than 1,000 PapA molecules assembled in a right-handed helical manner, with 3.3 molecules per turn [6,10].

All pilin subunits adopt an Ig-like fold but lack the seventh, C-terminal G β-strand, thus producing a large hydrophobic groove on the side of the protein (Figure 1 and [11,12]). In a process called donor–strand complementation (DSC), the G1 β-strand of PapD inserts a conserved motif of three alternating hydrophobic residues (called the P1 to P5 residues) plus N101 (P4 residue) into four binding pockets in the hydrophobic groove of the pilus subunits (P1 to P4 binding pockets). The G1 strand provides the structural information lacking in the pilus subunit by completing its Ig fold [11–13]. Pilus subunit assembly proceeds through a noncovalent polymerization process called donor–strand exchange (DSE; Figure 1). All subunits, except for the adhesin, possess an N-terminal extension (Nte) peptide of 11 (PapK), 12 (PapE and PapF), 19 (PapA), or 33 (PapH) residues (Figures 1 and S1) that is disordered and not part of their Ig-like structure. The Nte comprises a highly conserved array of alternating hydrophobic residues, called the P2 to P5 residues [14,15]. This array constitutes the DSE region of the Nte (see Figure S1 for location of this DSE region). As chaperone–subunit complexes are differentially targeted to the usher [16,17], each subunit donates its Nte to complete the Ig-fold of the subunit previously assembled by inserting its P2–P5 residues into the corresponding P2–P5 binding pockets, thus first displacing and then replacing the chaperone G1 strand in the groove of the previously assembled pilus subunit [18–21]. This process occurs through a zip-in–zip-out process whereby the DSE reaction is initiated by the insertion of the P5 residue of the Nte of one subunit into the P5 pocket of the groove of the other [21]. This binding event leads to the formation of a transient ternary complex, the formation of which is essential for the DSE reaction to proceed. Insertion of the Nte of PapH into the groove of a PapA subunit terminates pilus biogenesis.

Author Summary

Bacterial adhesion to a host is a crucial step that determines the onset of bacterial infection. It is mediated through recognition of a receptor on the host cell surface by a protein called an adhesin displayed on the surface of the bacterium. Many adhesins are displayed at the tip of specialized organelles called pili, some of which are assembled by the ubiquitous chaperone–usher pathway. In this pathway, each pilus subunit is assisted in folding by a chaperone. The resulting chaperone–subunit complex is targeted to a pore located in the outer membrane, called the usher, that serves as assembly platform. There, pilus subunits dissociate from the chaperone and polymerize, resulting in a surface organelle, the pilus, that protrudes out of the usher. Here, we have determined the structure of the major subunit of the P pilus, PapA. The P pilus, produced in uropathogenic Escherichia coli, displays the adhesin PapG responsible for targeting the bacterium to the kidney epithelium. We have determined the structure of PapA either bound to its cognate chaperone, PapD, or bound to another PapA subunit. These structures provide a view of PapA before and after its assembly in the pilus and shed light on the mechanism of PapA assembly.

Results/Discussion

Crystal Structures of PapD/PapANtd1_G15N and PapD/(PapANtd1_G15N_T101L)2

In order to investigate the PapD/PapA and PapA/PapA interactions within the P pilus rod, the first structure which we solved was that of PapD/PapA, where two mutations were introduced in the papA gene. This was necessary because PapA in purified wild-type PapD/PapA complexes tends to spontaneously polymerize, and self-polymerization of PapA has prevented structural studies. As PapA, like any other pilus subunits, polymerizes through DSE, mutating residues in the Nte of PapA before and after DSE. In that respect, the PapD/PapA Ntd1_G15N structure is not very informative. However, by investigating a different PapA mutant with the additional mutation T101L, we obtained crystals of a complex containing one PapD molecule and two PapA subunits. In this ternary complex, PapD is bound to PapA through DSC, and this subunit is itself bound to another PapA subunit through DSE. These structures are used as a basis for a detailed mutational study dissecting the requirements for PapA polymer formation.

Comparison between the Two PapA Molecules in the PapD/(PapANtd1_G15N_T101L)2 Structure

Figure 3A shows a superimposition of the two PapA molecules in the PapD/(PapANtd1_G15N_T101L)2 complex. With the donor–strand-complemented PapA subunit (residues 2 to 19; PapANtd1_G15N_T101L)2 complex is similar to the one obtained by Zavialov et al. for the Cag system [19]. designed that conserved the entire Nte but introduced an Asn at position 15. Position 15, a Gly residue in the wild-type Nte of PapA, locates in the DSE region of the Nte and is strictly conserved among all Pest Ntes (Figure S1). G15 is indeed required because, after DSE, it lies in the P4 pocket of the subunit’s grooves, which, in this region and in all Pest subunits, contains a bulky phenylalanine or a tyrosine (F152 in PapA, Y146 in PapK, F138 in PapE, F137 in PapF, and Y162 in PapH) [11,18]. We thus mutated G15 to N; however, PapAG15N in complex with PapD also undergoes spontaneous polymerization (Figure S2A, top panel). Thus, in addition to the G15N mutation, a mutant where residues 2 to 8 were deleted was next constructed. The deleted region is just N-terminal to the DSE region of the Nte of PapA. This mutant (PapANtd1_G15N) could not undergo spontaneous polymerization, and formed a stable complex with PapD (Figure S2A, middle panel). This experiment indicates that, as suggested by the electron microscopy study of Mu et al. [22], the region of the Nte N-terminal to the DSE region is involved in PapA/PapA interaction. Crystals of PapD/PapANtd1_G15N diffracted to a resolution of 2.6 Å, and the structure was solved by molecular replacement using the PapD/PapK structure as a search model. This structure is very similar to the already known PapD/PapK, PapD/PapENtd, or PapD/PapH Ntd1 structures [9,11,18]. Like PapK, PapE, or PapH, PapA lacks strand G of its Ig-fold; PapD complements this by donating its G1 strand.

By itself, the PapD/PapANtd1_G15N structure is not very informative. However, by investigating a different PapA mutant with the additional mutation T101L, we obtained crystals of a complex containing one PapD molecule and two PapANtd1_G15N_T101L molecules. This T101L mutant was initially designed to fill in the PapA P5 pocket. Indeed, as explained in the Introduction, all Pest subunits, except PapH, have a clear P5 pocket, which serves as an initiator point, the occupation of which triggers the DSE reaction [9,21]. Thus, the PapANtd1_G15N_T101L was made to test the possibility that by filling its P5 pocket, PapA would become more like PapH in being unable to undergo DSE. As shown in Figure S2B, indeed, PapANtd1_G15N_T101L has reduced DSE activity compared with that of PapANtd1_G15N. However, this T101L mutation had the additional unexpected consequence of stabilizing a complex containing a 1:2 molar ratio of PapD and PapA (Figure S2A, lower panel). PapD/(PapANtd1_G15N_T101L)2 was thus purified. Crystals were produced diffracting to 2.5 Å resolution. The structure of this ternary complex is shown in Figure 2. It clearly shows one PapA molecule bound to PapD through DSC in a complex very similar to PapD/PapANtd1_G15N; however, this time, the Nte of the donor–strand-complemented PapA molecule is bound to the groove of another PapA molecule, and thus this ternary complex crystal structure provides a snapshot of PapA before and after DSE. In that respect, the PapD/(PapANtd1_G15N_T101L)2 complex is similar to the one obtained by Zavialov et al. for the Cag system [19].

Recently, in a departure from the more conventional model described above, Mu et al. [22] suggested, based on electron microscopy and image reconstruction of the PapA rod, that the DSE region of the Nte of PapA is not involved in DSE, but instead the region of the Nte N-terminal to the DSE region is involved in the process. Interestingly, the Nte of PapA is longer than the Nte of most Pest subunits except PapH (Figure S1), and thus the model proposed by Mu et al. [22] could potentially explain why residues N-terminal to the DSE region would be required in the process of P pilus biogenesis. This is explored further in this report, in which we describe the structures of a binary complex of PapD bound to PapA and of a ternary complex containing the chaperone PapD and two Pest subunits. In this ternary complex, PapD is bound to PapA through DSC, and this subunit is itself bound to another Pest subunit through DSE. These structures are used as a basis for a detailed mutational study dissecting the requirements for PapA polymer formation.

Results/Discussion

Crystal Structures of PapD/PapANtd1_G15N and PapD/(PapANtd1_G15N_T101L)2

Comparison between the Two PapA Molecules in the PapD/(PapANtd1_G15N_T101L)2 Structure

Figure 3A shows a superimposition of the two PapA molecules in the PapD/(PapANtd1_G15N_T101L)2 complex, with the donor–strand-complemented Pest subunit (residues 2 to 19; PapANtd1_G15N_T101L)2 complex is similar to the one obtained by Zavialov et al. for the Cag system [19].
subunit (dsePapA) in orange. The core sheet structure of dsePapA is in a closer conformation than that of dscPapA, as the β-strands on each side of the groove of dsePapA are nearer to each other. Also, the "63–74" loop is ordered in dsePapA and not in dscPapA, as this molecule is missing residues 70 to 73 in this region. The truncated Nte of dscPapA (as indicated above, residues 2 to 8 were removed to create PapANtd1) is clearly visible in the groove of dsePapA from residue 10 (the two first residues of PapANtd1_G15N_T101L, which are residues 1 and 9 of full-length PapA, were not defined in the electron density). DsePapA is only visible in the electron density from residue 20, as its Nte is not interacting in the groove of another PapA molecule and is thus disordered. Figure 3B shows the surface of dsePapA bound to the truncated Nte of dscPapA (left panel) and that of dscPapA bound to the G1 strand of the chaperone (right panel): the PapD G1 strand interacts as expected at the P1 to P4 positions in the groove of dscPapA (right panel), the Nte of which interacts in the P2 to P5 pockets in the groove of dsePapA (left panel). It is noticeable in the left panel of Figure 3B that the dsePapA groove is extending beyond the region occupied by the P2 residue of dscPapA Nte. However, the groove of dscPapA is not extending beyond the P1 pocket, due to the disordered "63–74" loop. Modeling of the first nine residues of PapA Nte in the extended groove of dsePapA shows that this extended groove has the right length and shape to accommodate the nine missing residues in the Nte of PapANtd1 (unpublished data). Thus, the groove of PapA is long enough to accommodate the extended Nte of another PapA molecule.

Figure 1. Schematic Diagram Illustrating the Processes of DSC and DSE

In this diagram, β-strands and helices are represented as arrows and cylinders, respectively. At left, a topology diagram of a subunit bound to the chaperone is shown in pink. The Nte is shown as a disordered region at the N-terminus of strand A1. In the chaperone, only the G1 strand is shown in gray; it complements the fold of the subunit by inserting between strands A2 and F, providing in trans the missing G strand. Note that the complementing G1 strand runs parallel to strand F, thereby reconstituting an atypical Ig-fold for the subunit. The donor-strand-complemented subunit is referred to in the text as "dsc". At right, the same subunit in pink is shown in the pilus. Its Nte is now inserted between strand A2 and F of the subunit in green. Its groove is now filled with the Nte of the subsequent subunit in dark blue. The subunit is referred to in the text as "dse".

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Structure Comparisons of the Grooves of dscPapA in PapD/ PapA<sub>Ntd1_G15N</sub> and dsePapA in PapD/(PapA<sub>Ntd1_G15N_T101L</sub>)<sub>2</sub>

Figure 4A shows details of the grooves of dscPap<sub>A</sub> in PapD/ PapA<sub>Ntd1_G15N</sub> (left panel) and dsePap<sub>A</sub> in PapD/(PapA<sub>Ntd1_G15N_T101L</sub>)<sub>2</sub> (right panel). Figure 4A on the left panel shows that in the PapD/ PapA<sub>Ntd1_G15N</sub> structure, the P<sub>4</sub> position of the groove is formed by F<sub>152</sub>. This configuration of F<sub>152</sub> is similar to that seen in the equivalent position of the PapD/PapK, PapD/PapE, PapD/PapH, and PapE/K<sub>Nte</sub> complexes [9,11,18]. As mentioned above, the bulk created in the P<sub>4</sub> pocket by F<sub>152</sub> and equivalents, by being able to accommodate only a conserved Gly in the Ntes, acts as a registering device that calibrates the positioning of Ntes in the subunits’ grooves.

In contrast to what is observed in the PapE/K<sub>Nte</sub> or any other chaperone–subunit complex structures, the side chain of F<sub>152</sub> has moved out of the P<sub>4</sub> pocket of the DsePapA structure (Figure 4A, right panel) and the P<sub>4</sub> pocket can now accommodate the N<sub>15</sub> mutation (substituted for wild-type G<sub>15</sub> in the PapA<sub>Ntd1_G15N_T101L</sub> mutant to prevent higher-order polymerization). F<sub>152</sub> in the dsePapA structure is allowed to move away from the groove position because of the T<sub>101L</sub> mutation. Indeed, as shown in Figure 4B, the T<sub>101L</sub> mutation and the insertion of N<sub>15</sub> induce a rearrangement of the side-chains in the P<sub>4</sub>–P<sub>5</sub> region, leading to T<sub>99</sub> moving out of the P<sub>5</sub> region, thereby leaving room for F<sub>152</sub> to substitute in its place, the new F<sub>152</sub> position being stabilized by L<sub>101</sub>.

Structure Comparisons between the Groove of dsePapA in the PapD/ (PapA<sub>Ntd1_G15N_T101L</sub>)<sub>2</sub> Structure and That of dsePepE in the PapE/K<sub>Nte</sub> Structure

Like dsePapA in the PapD/(PapA<sub>Ntd1_G15N_T101L</sub>)<sub>2</sub> complex structure, PapE in the PapE/K<sub>Nte</sub> structure is also donor–strand exchanged (dsePepE), but with the Nte of PapK (K<sub>Nte</sub>) [18]. Comparing dsePepE (Figure 5A, right) with dsePapA (Figure 5A, left) shows that, while the groove of dsePepE stops at the P<sub>2</sub> pocket, the groove of dsePapA is extending beyond this pocket. This is due to the presence of the “63–74” loop in dsePapA, a loop which is much shorter in dsePepE, and to the closure of dsePepE groove by PapE N- and C-termini compared with the open groove in dsePapA (Figure 5B).
A number of PapA Nte mutants were next produced in order to evaluate the effect of these mutations on polymerization and pilus formation. In addition to PapA wild-type, six constructs were studied. In the DSE region, a G15N mutation and a D11–17 deletion (where the entire DSE region is removed) were made. In the region N-terminal to the DSE region of the Nte, an I4G single-site mutant and the Ntd1 deletion described above were studied. I4 is a bulky residue in that region of the Nte and thus would contribute significantly to the interface, were it to be involved in groove/Nte interaction (see [21] for consideration regarding surface area contribution of residues in the Nte). We also combined the Ntd1 (Δ2–8) deletion with the G15N mutation and the I4G and G15N mutations. Polymerization was assessed by gel filtration immediately after purification of the corresponding PapD/PapA complexes, and pilus formation was assessed after freeze–thaw of PapD/PapA complex preparations using electron microscopy (EM). Thus, gel filtration provides information on the limited polymerization events taking place early on during polymerization, while freeze–thaw of PapD/PapA complexes followed by analysis by EM provides information on the ability of the various PapA molecules to form pili. Results obtained for each of the PapA constructs are presented in Figure 6, where each panel provides the elution profile and EM micrograph for each wild-type and mutated PapA. For those complexes that exhibited a gel filtration peak corresponding to PapD/(PapA)2, further polymerization was checked by pooling the fractions corresponding to PapD/(PapA)2 and running another gel filtration the next day. For those complexes that did not exhibit polymerization, the PapD/(PapA)1 peak was rerun on gel filtration to make sure that indeed no polymers were formed.

Gel filtration of PapD/PapA wt complexes reveals a major 1:2 PapD/(PapA) complex. Yet, this complex polymerizes, as (1) rerunning a gel filtration on the 1:2 complex 24 hours later results in higher order polymers being formed (unpublished data), and (2) pili are formed readily (Figure 6A). These results confirm the existence of a rate-limiting step in the DSE reaction whereby PapD/(PapA)2 complex formation appears to be required before DSE can proceed to higher-order polymer forms [23]. The reason for such a rate-limiting step is unclear; it may be that this provides a checkpoint mechanism before committing to full biogenesis of the rod. The PapD/PapAwt behaves very much like the PapD/PapAwt in the gel filtration, but fewer pili appear to be made (Figure 6B). The single G15N mutation in the DSE region appears to affect polymerization with the detection of 1:3 and 1:4 PapD/PapA complexes (PapD/(PapA)3 and PapD/(PapA)4), and while pilus formation does occur, the diameters of the pili and their central channels are increased (Figure 6C). Thus, these mutations appear to slow down the reaction in such a way that polymer intermediates are observed after 24 h, but overall, these mutations do not seem to affect the process so stringently that it is unable to proceed.
PapANtd1 appears to be able to form 1:2 PapD/PapA complexes and aggregates of protein are visible by EM, the PapD/PapA Ntd1 mutant is totally impaired. Thus, both the DSE region and the region N-terminal to it are important for pilus formation. Site-directed and deletion mutagenesis confirm this view and thus validate earlier published observations by Mu et al. [22], which emphasized that both the DSE region and the region N-terminal to it are important for pilus formation. Site-directed and deletion mutagenesis confirm this view and thus validate earlier published observations by Mu et al. [22], which emphasized the role of the non-DSE region of the Nte. Thus, PapA uses an extended Nte and in that respect appears to be very similar to other major pilus subunits such as SafA of Salmonella or Caf1 of Yersinia [19,21]. The Nte of SafA, for example, is 17 residues long and one residue outside its DSE region, F3, was shown to be important in capping the process of DSE and driving the reaction to completion (the DSE region of SafA consists of residues 11 to 17). Indeed, a mutation of F3 to Ala in SafA results in an equilibrium between reaction species because in this mutant, DSE is allowed to proceed in reverse. A possible equivalent of F3 in PapA is I4. Indeed, DSE is somewhat affected by the I4G mutation. Thus, some features common to the assembly of all major pilus subunits are emerging, which include the involvement of an extended protein–protein interface and a potential capping mechanism driving polymer formation to completion. The PapA polymer is however different from the SafA or Caf1 polymer in that it adopts a distinct tertiary superhelical structure. The structures presented here do not provide any clues as to how such a ternary structure could form. Indeed, packing interfaces observed in both the PapD/PapANtd1_G15N and PapD/ (PapANtd1_G15N_T101L)2 crystals appear irrelevant (unpublished data). However, the structure of PapA elucidated here...
provides the basis for complementing the work by Mu et al. [24] and characterizing further the PapA/PapA interactions that preside over superhelix formation.

**Materials and Methods**

**Plasmid preparations.** See Text S1.

**Protein expression and purification of the PapDHis/PapA complexes.** The eight PapDHis/PapA constructs (PapDHis/PapAwt, PapDHis/PapANtd1, PapDHis/PapAD11–17, PapDHis/PapAG15N, PapDHis/PapAI4G, PapDHis/PapANtd1_G15N, PapDHis/PapAI4G_G15N, and PapDHis/PapANtd1_G15N_T101L) were transformed one at a time into *E. coli* C600 cells and grown in a 5-l fermentor vessel containing Terrific Broth (TB; Sigma, http://www.sigmaaldrich.com) kept at 37°C and shaken at 600 rpm. The cells were induced with 1 mM IPTG, once the OD600 reached a value of 0.9, and kept growing for another 3.5 h. The complexes were purified after periplasmic extraction using Cobalt-affinity chromatography (Talon; Clontech, http://www.clontech.com), followed by hydrophobic interaction chromatography (phenyl source; GE Healthcare, http://www.gehealthcare.com), ending with a gel filtration step in 20 mM Tris-HCl (pH 7.5) and 20 mM NaCl, using a Superdex75 120 ml column. This last step was crucial for separating the different polymer forms of PapA in complex with PapD (PapD/(PapA)1, PapD/(PapA)2, ...). The Purification of PapDHis/(PapANtd1_G15N_T101L)2 complex was purified as explained above. However, after the gel filtration step, there was a second major peak that eluted at a volume of around 54 ml (the 1:1 complex eluted at around 60 ml) and was interpreted as a 1:2 complex (PapD/(PapA)2). The 1:2 complex was concentrated to 13 mg/ml for crystallization trials.

**Purification of PapD/PapANtd1_G15N.** The PapD/PapANtd1_G15N complex was purified from periplasmic extracts using cation-exchange chromatography (SP HiTrap HP column; GE Healthcare) followed by hydrophobic interaction chromatography (phenyl source). The purification was completed by a gel filtration step in 20 mM MES (pH 6.0) and 20 mM NaCl on a Superdex75 120 ml column. The 1:1 complex eluted at a volume of around 60 ml and was concentrated to 8 mg/ml for crystallization trials.

**In vitro DSE experiments.** See Text S1.

**PapD/PapANtd1_G15N and PapDHis/(PapANtd1_G15N_T101L)2 crystallization and structure determination.** PapD/PapANtd1_G15N was crystallized at room temperature in a hanging drop. In the first crystal form (plates), the drop was equilibrated against a reservoir solution containing 25% PEG8K, 10% isopropanol, and 0.1 M MES (pH 6.6). In the second crystal form (rods), the drop was equilibrated against a reservoir solution containing 2 M ammonium sulfate and 0.1 M Na acetate (pH 5.6). The plates belonged to space group C2 and diffracted to 3.2 Å, whereas the rods belonged to space group P321 with cell dimensions a = 167 Å, b = 167 Å, c = 178 Å, and diffracted to 2.6 Å. The solvent content is 71%, with 4 PapD/PapANtd1_G15N complexes per asymmetric unit (Table S2). The structure solved by molecular replacement using PapD/PapK as a search model with the program AMoRe [25]. The PapK molecule was first modeled to a poly-alanine chain prior to refinement. The first refinements were performed using simulated annealing with CNS and simulated annealing with ARP/wARP [26].

**Figure 6.** Gel Filtration Profiles and EM pictures of PapD/PapA Wild-Type (wt) and Mutant Complexes

Each gel filtration profile of a particular PapA construct is shown with a view of its corresponding EM picture on the left. The complex is labeled on the right. Only the gel filtration peaks corresponding to free PapD, PapD/(PapA)1 (1/1) or PapD/(PapA)2 (1/2) are labeled. Representative pili are labeled with arrows (A–C). Non-pilus polymerization of protein is labeled with arrowheads (D) and (F). The scale of all the EM pictures is represented by the black bar on the PapD/PapAwt picture and corresponds to 100 Å.

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annealing and noncrystalllographic symmetry restraints for the four complexes in the asymmetric unit, using CNS [26]. Then successive cycles of manual rebuilding with O [27] and conjugate gradient minimization with CNS were performed. B factors were refined individually. Toward the end of the refinement, the noncrystalllographic symmetry restraints were only applied to some parts of the β-sheets of the complexes. The refinement converged to the final R values of R = 23.0% and Rfree = 26.6% with good stereochemistry. PapD/PapAcomplexes (PDB codes 2uy6, 2uy7, 2uyl, 2uy5) were crystallized at 10–20 mM TrisHCl (pH 7.6) and 20–50 mM NaCl. The frozen–thawed sample (5 μl) was placed on carbon-coated, glow-discharged grids, washed with 10 mM TrisHCl (pH 7.6), negatively stained with 1% uranyl acetate, and imaged on a Philips CM12 electron microscope. For each sample, 109 plates were frozen and thawed five times in liquid nitrogen at 150 g. Then successive cycles of conjugate gradient minimization with CNS were performed. B factors were refined individually. Toward the end of the refinement, the noncrystallographic symmetry restraint was applied between the two PapA molecules. The refinement converged to the final values of R = 22.3% and Rfree = 26.2% with good stereochemistry (Table S2).

Electron Microscopy of the PapD/PapA Complexes. PapD/PapA (chaperone/pilin) samples of wild-type and mutant PapA proteins were frozen and thawed five times in liquid nitrogen at 150 μg/ml in 10–20 mM TrisHCl (pH 7.6) and 20–50 mM NaCl. The frozen–thawed sample was used either on the same day, or allowed to sit at 4°C for as long as 40 d to enhance oligomerization of PapA subunits. The sample (5 μl) was placed on carbon-coated, glow-discharged grids, washed with 10 mM TrisHCl (pH 7.6), negatively stained with 1% uranyl acetate, and imaged on a Philips CM12 electron microscope (no longer available).

Supporting Information

Figure S1. Sequence Alignment of Pap Subunit Ntes

The sequences have been aligned using the conserved Glycine (pink boxes) and the alternating hydrophobic residues of the DSE region (indicated). The P2 to P5 residues (yellow boxes) of the Nte that insert into the P2 to P5 regions (or pocket) of the subunit grooves are indicated. The C-terminal borders of the Ntd1 and Δ11–17 deletions are indicated.

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